

## Supplementary Information

### Conditional Antisense Oligonucleotide Triggered by miRNA

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Figure S1. NUPACK calculation of free energy ( $\Delta G$ ) for **conditional EZN2968** at 37 °C.

Figure S2. miR-122 expression levels in different cell lines.

Figure S3. Spontaneous activation of **conditional EZN2968** in U373 cells.

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Figure S5. Activation kinetics of **T<sub>7</sub>B<sub>3</sub>\*** incubated with miR-122, scr. 1-7nt miR-122 or scr. miR-122.

Figure S6. Fluorescence lifetime measurement to evaluate specificity of **T<sub>7</sub>B<sub>3</sub>\*** to miR-122 in buffer.

Figure S7. Endogenous miR-122 in Huh7 cells can be knocked down using anti-miR-122.

Figure S8. miR-122 and miR-21 expression levels in U373 cells.

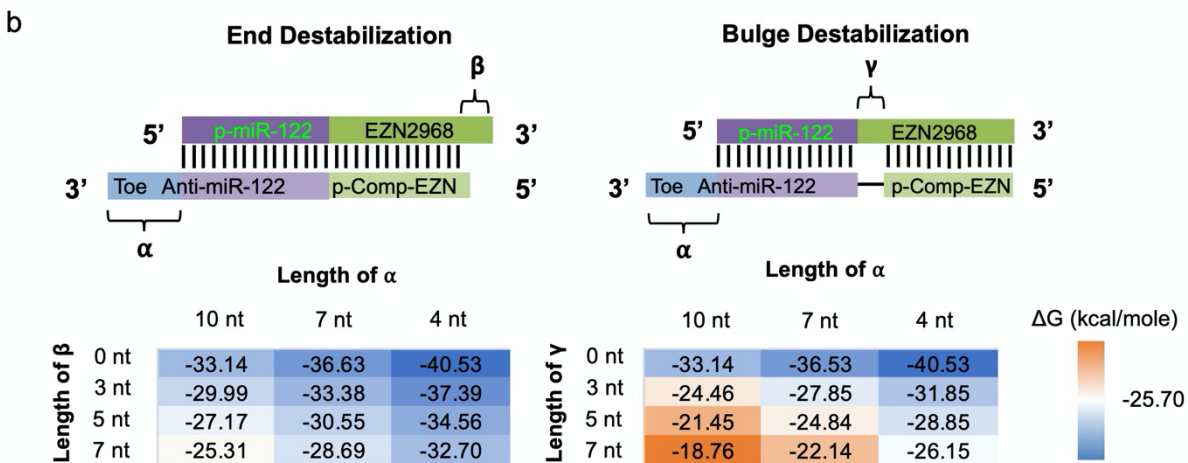
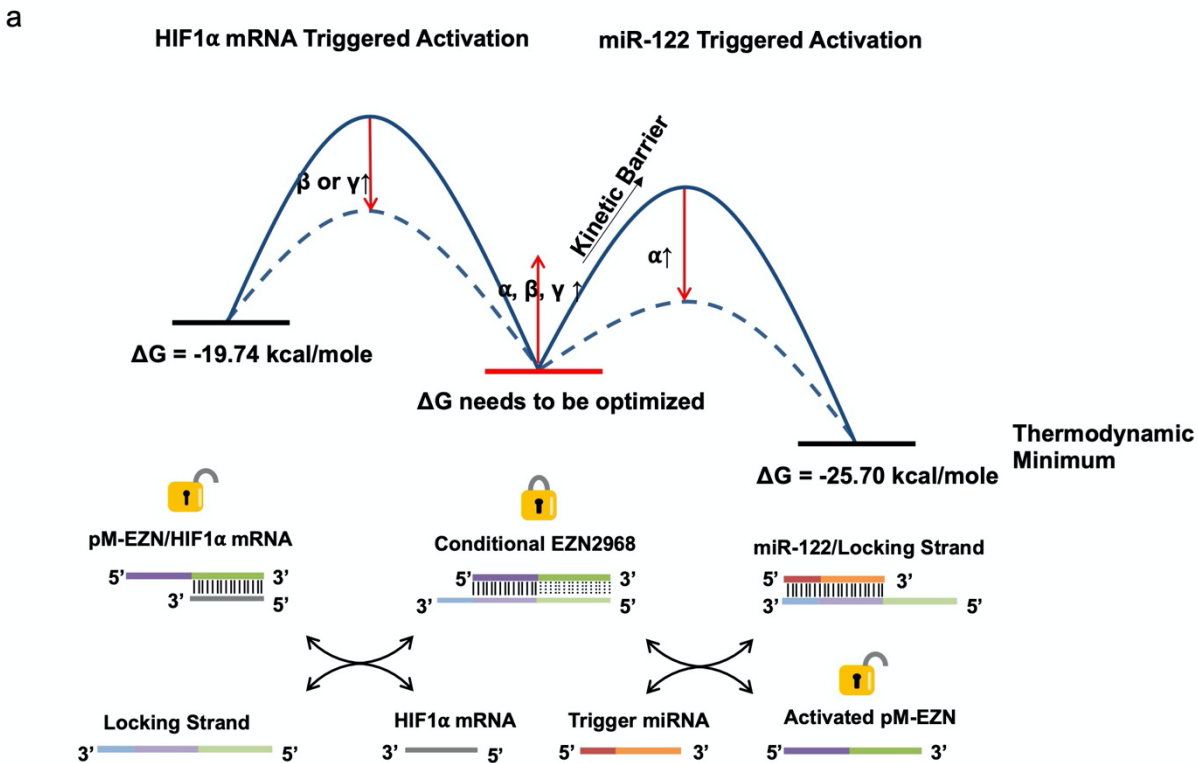
Figure S9. Flow cytometry of U373 cells transfected with miR-21 or miR-122 inducible **conditional EZN2968**.

Table S1. Oligonucleotide sequences for in buffer testing of leakage activation of bulge and end destabilized **conditional EZN2968**.

Table S2. Oligonucleotide sequences for *in vitro* testing of **conditional EZN2968**.

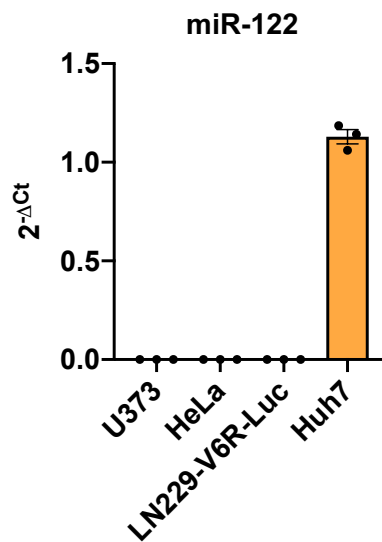
Table S3. Oligonucleotide sequences for miR-21-inducible EZN2968.

Table S4. Primer Sequences.

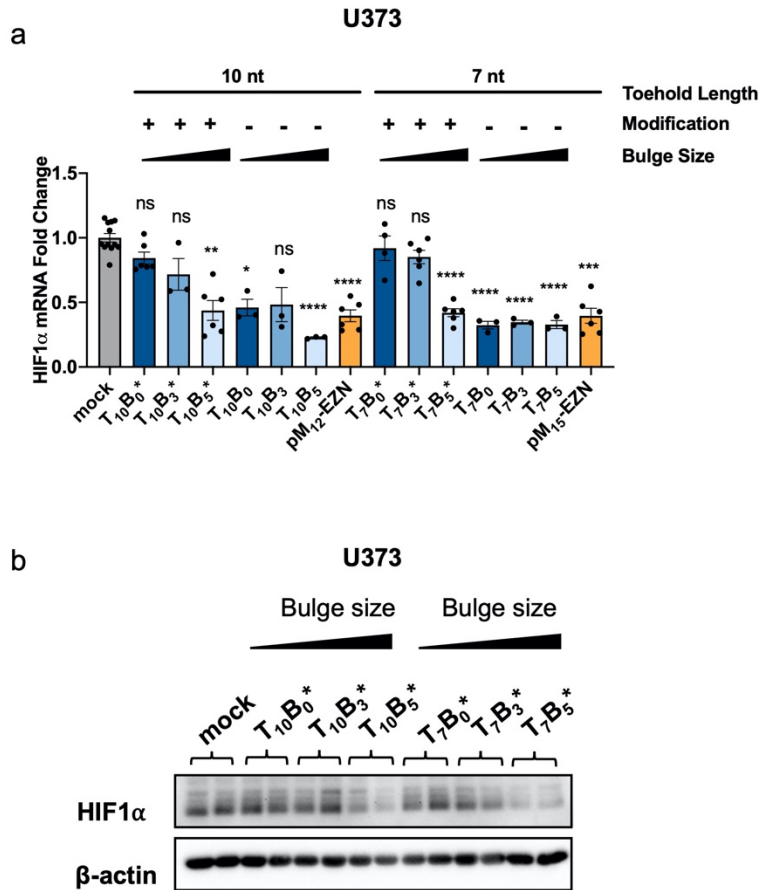


**Figure S1. NUPACK prediction of free energy ( $\Delta G$ ) for conditional EZ2968 at 37 °C.** (a) Energy diagram showing displacement of the pM-EZN strand by miR-122 or displacement of the locking strand by HIF1 $\alpha$  mRNA. The predicted  $\Delta G$  is -25.70 kcal/mole for miR-122/locking strand duplex, and -19.74 kcal/mole for pM-EZN/HIF1 $\alpha$  mRNA. The calculation indicates that the conditional ASO binding to miRNA is more favorable than binding to the target mRNA. This design minimizes mRNA-mediated unlocking of the conditional ASO. The toehold length ( $\alpha$ ,  $\beta$  and  $\gamma$  domains) cannot be independently tuned without also tuning the stability of the conditional ASO. Increasing the length of the  $\beta$  or  $\gamma$  domain destabilizes the conditional ASO and also reduces

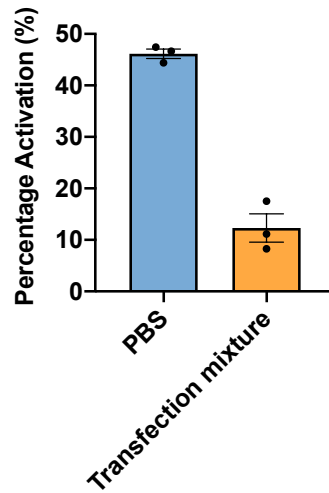
the barrier to binding the mRNA target. Likewise, increasing the length of the  $\alpha$  domain destabilizes the conditional ASO and lower the barrier to miRNA binding. (b) Predicted  $\Delta G$  of conditional ASO duplexes with different length of  $\alpha$ ,  $\beta$  and  $\gamma$  domain. When  $\Delta G > -25.70$  kcal/mole, miRNA triggered activation is favorable. These values were calculated using the default parameter set for DNA (SantaLucia, 1998) in NUPACK with 0.15 M NaCl and 10 nM oligonucleotide concentrations at 37°C.



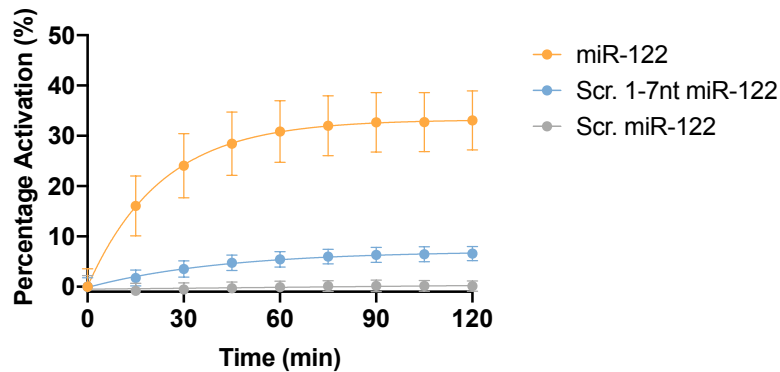
**Figure S2. miR-122 expression levels in different cell lines.** RNA was isolated from each cell line using miRNeasy Mini Kit (QIAGEN), and reverse-transcribed using miScript II RT Kit (QIAGEN). miR-122 levels were quantified by qPCR with RNU6 as a reference.  $\Delta Ct = Ct_{miR-122} - Ct_{RNU6}$ . The error bars represent SEM from triplicate measurements.



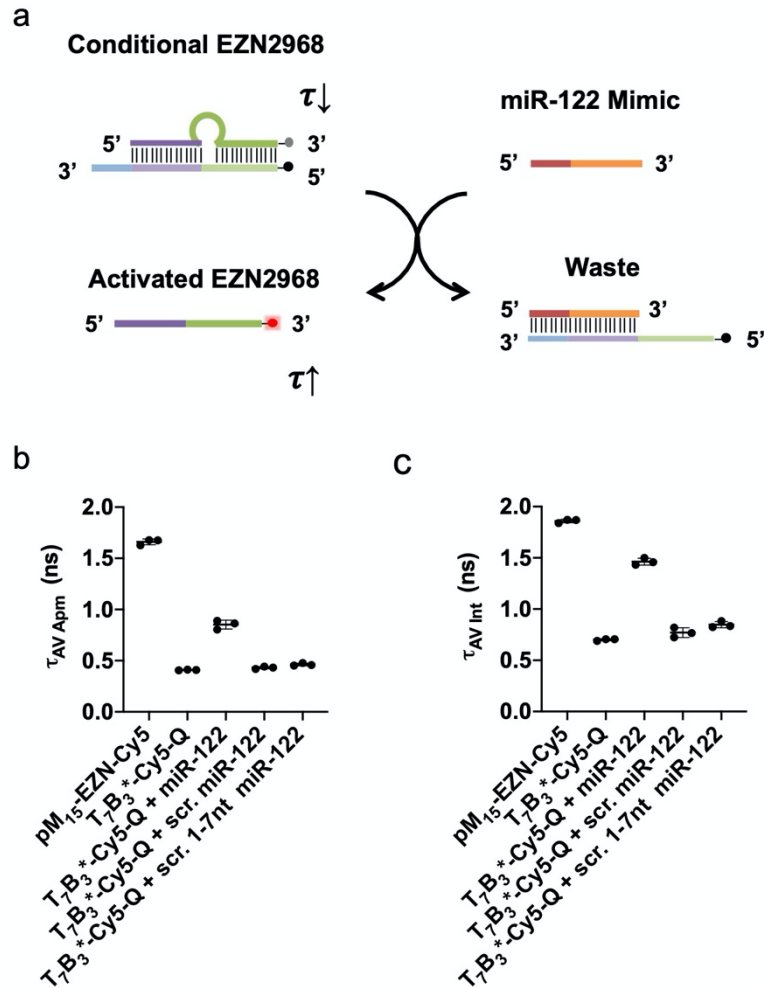
**Figure S3. Spontaneous activation of conditional EZN2968 in U373 cells.** U373 cells were transfected with 10 nM conditional EZN2968 using Oligofectamine and incubated for 24 h. (a) HIF1 $\alpha$  mRNA levels were quantified by qPCR normalized to 18S. The error bars represent SEM from  $n =$  or  $> 3$  measurements. \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , \*\*\*\*  $p < 0.0001$ , Brown-Forsythe and Welch ANOVA tests with Dunnett's T3 multiple comparison. (b) Western blot of HIF1 $\alpha$  protein with  $\beta$ -actin used as a housekeeping gene.



**Figure S4. Conditional EZN2968 activation in transfection mixture.** 10 nM Cy5/quencher-labeled  $T_7B_3^*$  were incubated with 500 nM miR-122 mimic in PBS or in transfection mixture. For the transfection mixture group, Cy5/quencher-labeled  $T_7B_3^*$  and miR-122 mimic were complexed with Oligofectamine in OptiMEM first and then mixed together, using the same procedure as the *in vitro* transfection experiment.

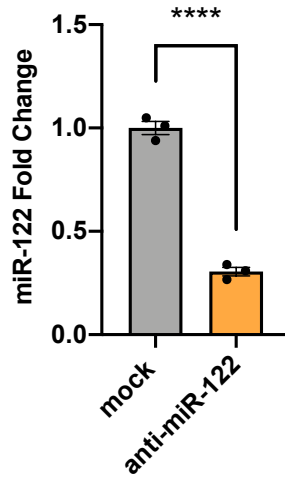


**Figure S5. Activation kinetics of T<sub>7</sub>B<sub>3</sub>\* incubated with miR-122, scr. 1-7nt miR-122 or scr. miR-122.** 10 nM Cy5/quencher-labeled T<sub>7</sub>B<sub>3</sub>\* and 100 nM miR-122, scr. 1-7nt miR-122, or scr. miR-122 were incubated at 37°C for 2h in a plate reader and fluorescence was measured with an interval of 15 min. Percentage activation was determined based on the fluorescence increase. 10 nM Cy5-labeled pM-EZN strand was used as a positive control, whose fluorescence intensity represents 100% activation.

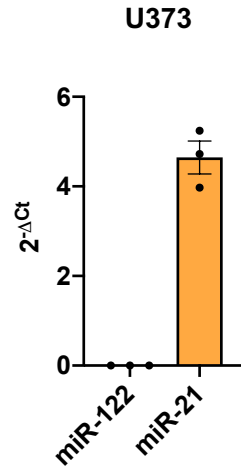


**Figure S6. Fluorescence lifetime measurement to evaluate specificity of  $T_7B_3^*$  to miR-122 in buffer.** (a) Scheme showing Cy5 fluorescence lifetime increase due to activation of  $T_7B_3^*$  by miR-122 mimic. (b) Amplitude-averaged and (c) intensity-averaged Cy5 fluorescence lifetime of 10 nM Cy5/Q-labeled  $T_7B_3^*$  solution incubated with 500 nM miR-122, scr. miR-122, or scr. 1-7 nt miR-122 for 2 h at 37 °C in PBS. Unlocked  $pM_{15}$ -EZN-Cy5 was used as a positive control. The error bars represent SD.

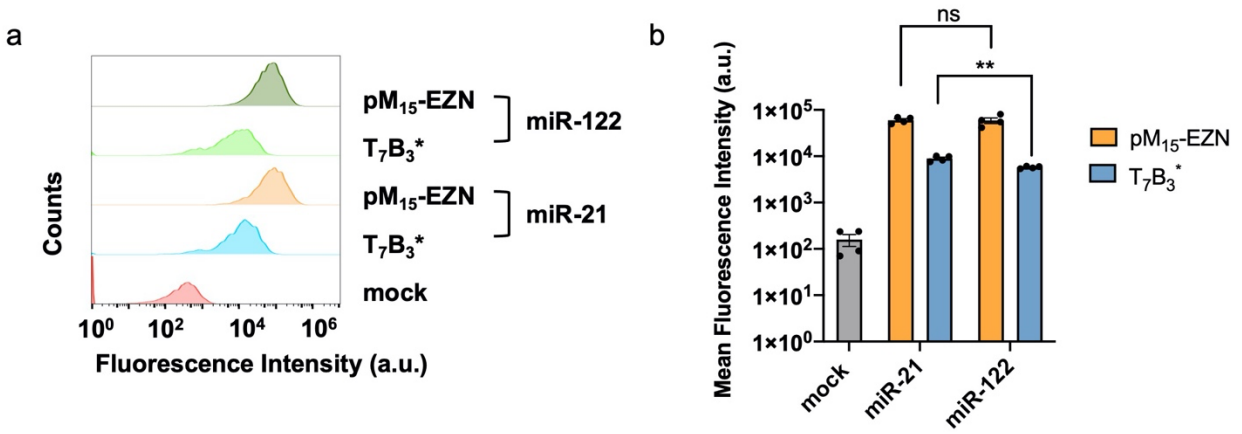




**Figure S7. Endogenous miR-122 in Huh7 cells can be knocked down using anti-miR-122.** Huh7 cells were transfected with 500 nM anti-miR-122 (Ambion) using Oligofectamine. After 24 h incubation, miR-122 levels were quantified by qPCR normalized to RNU6. \*\*\*\*  $p < 0.0001$ ,  $t$ -test.



**Figure S8. miR-122 and miR-21 expression levels in U373 cells.** RNA was isolated from U373 cells using miRNeasy Mini Kit (QIAGEN), and reverse-transcribed using miScript II RT Kit (QIAGEN). miRNA levels were quantified by qPCR with RNU6 as a reference.  $\Delta\text{Ct} = \text{Ct}_{\text{miR-122}} - \text{Ct}_{\text{RNU6}}$ . The error bars represent SEM.



**Figure S9. Flow cytometry of U373 cells transfected with miR-21 or miR-122 inducible conditional EZN2968.** (a) Histogram and (b) mean fluorescence intensity of U373 cells transfected with 10 nM Cy5/Q-labeled miR-21- or miR-122-inducible  $T_7B_3^*$  and incubated for 24 h. Cells transfected with 10 nM corresponding  $pM_{15}$ -EZN strands labeled with Cy5 were used as positive controls. The error bars represent SEM. \*\*  $p < 0.01$ ,  $t$ -test.

**Table S1.** Oligonucleotide sequences for in buffer testing of leakage activation of bulge and end destabilized conditional EZN2968

/3AmMO/= 3' amino modification; /5IAbRQ/=5' Iowa Black RQ; /5AmMC6/=5' amino modification; "T" indicates a mismatch in the toehold binding region.

ID	Sequence
Unmodified pM <sub>12</sub> -EZN	CAA TGG TGT TTG TGG CAA GCA TCC TGT A/3AmMO/
Unmodified pM <sub>15</sub> -EZN	TGA CAA TGG TGT TTG TGG CAA GCA TCC TGT A/3AmMO/
Unmodified pM <sub>18</sub> -EZN	GTG TGA CAA TGG TGT TTG TGG CAA GCA TCC TGT A/3AmMO/
Unmodified B <sub>0</sub>	/5IAbRQ/TA CAG GAT GCT TGC CAC AAA CAC CAT TGT CAC <u>TCT</u> CCA
Unmodified B <sub>3</sub>	/5IAbRQ/TA CAG GAT GCT TGC AAA CAC CAT TGT CAC <u>TCT</u> CCA
Unmodified B <sub>5</sub>	/5IAbRQ/TA CAG GAT GCT CAA ACA CCA TTG TCA <u>CTC</u> TCC A
Unmodified B <sub>7</sub>	/5IAbRQ/TA CAG GAT GCA AAC ACC ATT GTC <u>ACT</u> CTC CA
Unmodified E <sub>3</sub>	/5IAbRQ/AG GAT GCT TGC CAC AAA CAC CAT TGT CAC <u>TCT</u> CCA
Unmodified E <sub>5</sub>	/5IAbRQ/GA TGC TTG CCA CAA ACA CCA TTG TCA <u>CTC</u> TCC A
Unmodified E <sub>7</sub>	/5IAbRQ/TG CTT GCC ACA AAC ACC ATT GTC <u>ACT</u> CTC CA

**Table S2.** Oligonucleotide sequences for *in vitro* testing of conditional EZN2968

“+” = LNA modification; “\*” = PS modification; /3AmMO/ = 3’ amino modification; “I” indicates a mismatch on the toehold binding region.

ID	Sequence
EZN2968	+T*+G*+G* C*A*A* G*C*A* T*C*C* +T*+G*+T* A
EZN3088	+C*+G*+T* C*A*G* T*A*T* G*C*G* +A*A*+T* C
Modified pM <sub>12</sub> -EZN	C*A*A* T*G*G* T*G*T* T*T*G* +T*+G*+G* C*A*A* G*C*A* T*C*C* +T*+G*+T* A/3AmMO/
Modified pM <sub>15</sub> -EZN	T*G*A* C*A*A* T*G*G* T*G*T* T*T*G* +T*+G*+G* C*A*A* G*C*A* T*C*C* +T*+G*+T* A/3AmMO/
Modified B <sub>0</sub> (B <sub>0</sub> *)	/5IAbRQ/+T*+A* +C*A*G* G*A*T* G*C*T* T*G*C* C*A*C* A*A*A* C*A*C* C*A*T* T*G*+T* +C*A*C* A*C*T* C*C*A
Modified B <sub>3</sub> (B <sub>3</sub> *)	/5IAbRQ/*T*A* C*A*G* G*A*T* G*C*+T* +T*+G*+C* +A*A*A* C*A*C* C*A*T* T*G*T* C*A*C* A*C*T* C*C*A
Modified B <sub>5</sub> (B <sub>5</sub> *)	/5IAbRQ/*T*A* C*A*G* G*A*T* +G*+C*+T* +C*+A*+A* A*C*A* C*C*A* T*T*G* T*C*A* C* <u>I</u> *C* T*C*C* A
B <sub>3</sub> * without toehold	/5IAbRQ/T*A* C*A*G* G*A*T* G*C*+T* +T*+G*+C* +A*A*A* C*A*C* C*A*T* T*G*T* C*A
miR-122 mimic	+T*G*G* +A*G*T* +G*T*G* +A*C*A* +A*T*G* +G*T*G* +T*T*T* +G
Scrambled miR-122	+G*A*A*+G*T*A*+T*G*T*+G*G*T*+G*A*T*+T*G*C*+G*T* G*+T
Scr. 1-7nt miR-122	+G*G*G*+T*T*G*+A*T*G*+A*C*A*+A*T*G*+G*T*G*+T*T* T*+G

**Table S3.** Oligonucleotide sequences for miR-21-inducible EZN2968

“+” = LNA modification; “\*” = PS modification; /3AmMO/ = 3’ amino modification

<b>ID</b>	<b>Sequence</b>
pM <sub>15</sub> -EZN for miR-21-inducible EZN2968	T*C*A* G*A*C* T*G*A* T*G*T* T*G*A* +T*+G*+G* C*A*A* G*C*A* T*C*C* +T*+G*+T* A/3AmMO/
B <sub>3</sub> * for miR-21-inducible EZN2968	/5IAbRQ/*T*A* C*A*G* G*A*T* G*C*+T* +T*+G*+T* +C*+A*A* C*A*T* C*A*G* T*C*T* G*A*T* A*A*G* C*T*A

**Table S4.** Primer Sequences.

<b>Primer</b>	<b>Sequence (5'→3')</b>
HIF1 $\alpha$ Forward	TATGAGCCAGAAGAAGCTTTTAGGC
HIF1 $\alpha$ Reverse	CACCTCTTTTGGCAAGCATCCTG
18S Forward	AGGAATTGACGGAAGGGCACCA
18S Reverse	GTGCAGCCCCGGACATCTAAG